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sampling for leaf nutritional analysis - FCC

1. take a sample for each type of kiwifruit
example G3 - Hayward - Soreli, you cannot make a single analysis on different varieties, each variety represents a sample to be analyzed in the laboratory.

2. identify the plot of interest. you can also do one sample per plot if it is not larger than two hectares, otherwise it is advisable to divide the plot in two and do a double analysis

3. once the variety and plot to be analyzed have been identified, sampling is done diagonally across the rows, avoiding taking leaves on the perimeter plants. the choice of leaves to be analyzed should fall on mature young leaves taken at mid-shoot (therefore, apical leaves or those close to the trunk should be avoided)

4. the sample to be sent to the laboratory should consist of a minimum of 30 leaves (if the leaves are large) to a maximum of 50 leaves (if the leaves are small)

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Methods of sampling water for irrigation water analysis /chemical and bacteriological potability analysis ASCC/PCC/PMM

Material

1. Sterile 500 ml plastic bottle (with thiosulfate in case of chlorinator implantation) PMM
2. Chemical analysis bottle (plastic) PCC or ASCC
3. Portable refrigerator .

Sampling method

4. remove rubber hoses and plastic filters
5. run water for 5 minutes if used often
6. run the water for about 15 minutes if it has not been used for a long time close the faucet
7. sterilize the faucet with a flame (use gas lantern or cotton ball soaked in alcohol) and reopen the faucet, let the water run for another 2 minutes and flambé again, take the sample with the sterile bottle taking care to open and close it as quickly as possible
8. draw water for chemical analysis(ASCC), after rinsing the container with the same water to be analyzed

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Soil sampling methods PED/EAQ

Exclude areas from the area to be sampled that are abnormal in appearance (color, texture, skeleton, etc.) and agronomic history (crop diversity, treatments, fertilization, etc.)
Exclude plot edges near ditches and from headlands
It is essential that the sample consists of at least 20 elementary samples (sub-samples) taken at different points and carefully mixed.
Sampling points should be chosen by following a random path through the entire field.
Always remove the grass sward before sampling (0-5 cm).
For herbaceous crops, a soil layer between 5 and 30 cm should be taken; for tree crops, it is preferable to take two samples, one between 5 and 30 cm and the other between 30 and 60 cm, both to be sent to the laboratory.
In case of homogeneous soil, one soil sample should be taken for every 4-6 ha. In case of inhomogeneous soil, the area should be reduced depending on the degree of inhomogeneity.

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Method of sampling plant material (for phytopathological analysis). - FITO

The sampler should preferably take those plants showing the symptoms deemed abnormal at an early stage; the sample should contain both healthy and diseased parts.
It is very important to accompany the samples with all information regarding: - cultivar - rootstock - age and origin of the plant - occurrence and extent of symptoms - crops previously present - distribution of diseased plants in the field (isolated episodes, in patches, in rows, at the edges, etc.).
In all those cases where sample delivery cannot be done on the same day, it is of paramount importance to store it in a refrigerator at 4-5 °C.
Storage should not exceed 2-4 days. Never store in the freezer. Use closed plastic bags for storage; avoid wrapping samples with paper or cloth.
Contact the laboratory before delivering samples.